

American Chemical Science Journal 15(3): 1-12, 2016, Article no.ACSJ.19789 ISSN: 2249-0205



SCIENCEDOMAIN international www.sciencedomain.org

Proximate Analysis and Toxicological Studies of Polyalthia longifolia Seed Flour in Dietary Formulation of Albino Rats

Ibironke Adetolu Ajayi^{1*} and Emmanuel Nnamdi Ifedi¹

¹Department of Chemistry, Industrial Unit, Faculty of Science, University of Ibadan, Ibadan, Nigeria.

Authors' contributions

This work was carried out in collaboration between both authors. Authors IAA and ENI designed the study, performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Authors IAA and ENI managed the analyses of the study. Authors IAA and ENI managed the literature searches. Both authors read and approved the final manuscript.

Article Information

DOI: 10.9734/ACSJ/2016/19789 <u>Editor(s):</u> (1) Silvia Antonia Brandán, Inorganic Chemistry Institute, National University of Tucumán (UNT), Argentina. <u>Reviewers:</u> (1) Manjoosha Srivastava, CSIR- National Botanical Research Institute, Lucknow, Uttar Pradesh, India. (2) Iyekhoetin Matthew Omoruyi, Benson Idahosa University, Benin City, Edo State, Nigeria. Complete Peer review History: <u>http://sciencedomain.org/review-history/15303</u>

Original Research Article

Received 26th June 2015 Accepted 15th December 2015 Published 7th July 2016

ABSTRACT

Aim: A short term toxicological study of *Polyalthia longifolia* seed flour in dietary formulation of albino rats was conducted and investigated in order to determine its suitability as an additive in feed supplement and formulation.

Methodology: The effect of the dietary formulation on the physical appearance, feed intake and weight gain of the rats was determined. The proximate analysis, mineral composition, haematological and blood biochemistry were determined and calculated using standard methods of analysis.

Results: The proximate analysis revealed that the seed flour has high carbohydrate and moderate protein contents. The values of $59.66\pm0.19\%$ for carbohydrate, $12.40\pm0.25\%$ for protein, $11.90\pm0.04\%$ for crude fiber, 90.80% for dry matter were obtained for *P. longifolia* seed flour. The seed flour was found to contain high concentration of iron, manganese, zinc, copper and potassium. Other mineral present such as magnesium, calcium and sodium were very low. Nickel, chromium and lead were not present. The low Na/K ratio (0.03) obtained showed that the seed flour could

probably reduce high blood pressure. Proximate composition of the compounded feed both for the control and the test groups pointed out that there were significance differences in crude protein and crude fibre contents of the feeds compounded. Weekly monitoring of the rats showed good physical appearance and steady weight gain with no mortality recorded during the period of the experiment. The average weight gain per rat at the end of the experiment was shown to be 86.58 g for control rats and 90.00 g for test ones.

Conclusion: Haematological analysis of the rats in both groups showed that they were not anemic and there was no significant difference at $p \le 0.05$ in their blood biochemistry. There was no lesion found in the sections of the kidney, liver and heart of the rats. *P. longifolia* seed flour seemed to be a good supplement in livestock feed formulation.

Keywords: Polyalthia longifolia; chemical analysis; toxicological studies; albino rats.

1. INTRODUCTION

Food crops have occupied an important place in human nutrition as they remain the major sources of calories and proteins for a large proportion of the world population, particularly, in the developing countries [1]. In many of these countries, the supply of animal protein is inadequate to meet the protein needs of the rapidly growing population and the consumption of high amount of meat increases the risk of cardiovascular diseases and some types of cancer. This has necessitated contemporary research that is directed towards studying the food properties and potential utilization of protein from locally available food crops, most importantly from underutilized or neglected high protein oilseeds and legumes [2,3,4]. The nutritional value of any ingredient or feed can be evaluated by biological, chemical and physical scores [5]. Chemical score has proven to be a vital tool in food chemistry because it tends to assess the nutritional value based on the proximate composition of the food and feed which includes the protein, carbohydrate, lipid, moisture, ash and dietary fibre contents respectively. Plants are primary sources of carbohydrate and other nutrients used by human on daily basis. Their roots, leaves, fruits and seeds often provide food for humans [6]. Due to insufficient availability and high cost of animal protein, there has been a constant search for unconventional legumes and oils seed as new source of protein for use as functional supplements in food system [7].

P. longifolia (Sonn.), which belongs to the family of Annonaceae, is an evergreen plant commonly used as an ornamental street tree due to its effectiveness in combating noise pollution [8]. The genus *Polyalthia* includes about 120 species occurring mainly in Africa, South and South-Eastern Asia, Australia, and New Zealand. The bark has been reported to be used in the treatment of skin diseases, fever, hypertension, helmenthiasis and in a febrifuge [9]. Its aqueous extract also lowers the blood pressure and rate of respiration in experimental animals [10].

The hypoglycemic and anti-hyperglycemic activity of various solvent extracts of P. longifolia leaf extracts was evaluated in alloxan-induced experimental diabetes in rats. P. longifolia extracts and powder produced glucose lowering activity, which suggested then that the extracts did not modify any of the biochemical parameter significantly [11]. The antimicrobial activity of the leaves [12], stem [13] and seed diterpenoids [14] have been reported. The plant P. longifolia has been reported to contain alkaloids. clerodane di-terpines, quercetin, bulbocapnin, campesterol, diterpenes, and sesquiterpenoids which may contribute to the analgesic activity [15]. P. longifolia seed flour, regarded as an under-utilized plant product and with its significant biological and pharmacological activities can be used as additive in the diet formulation. Food additives are substances which are usually included in feeds in trace amounts to preserve its nutritional characteristics prior to feeding (antioxidants and mould inhibitors): facilitate ingredient dispersion or feed pelleting (emulsifiers, binders or stabilizers); promote growth (growth promoters, antibiotics and hormones); facilitate feed intake and acceptance (feeding stimulants and colorants) and also supply some essential nutrients such as minerals and vitamins [16]. Additives improve the value of certain foods nutritional and can make them more appealing by improving their taste, texture, consistency or colour while some modern synthetic preservatives/additives are controversial because they have been shown to cause respiratory or other health problems. Some studies point to synthetic preservatives and artificial coloring agents aggravating symptoms in those affected [17].

We are living in a world where maximum utilization of natural resources is the ultimate goal. Currently, these seeds are not eaten nor used for any industrial purposes in Nigeria. Information on the chemical and nutritional composition of P. longifolia seed flour is very scanty while previous workers in the Western part of Nigeria have given information on the phytochemical and fatty acid profile analysis of P. longifolia seed oil [18]. There is no reported data on effect of incorporating this seed into the food chain. This paper therefore, reports on the proximate analysis and toxicological study of Polyalthia longifolia seed flour on dietary formulation in albino rats. This would provide scientific data regarding their potential application in food processing as well as additive in food supplements.

2. MATERIALS AND METHODS

2.1 Materials

P. longifolia seeds used for this work were collected from Agbowo, Ibadan, Oyo State of Nigeria. The plant seeds were identified and authenticated at the Herbarium Unit of Botany Department, University of Ibadan, Nigeria.

2.2 Physical Characterization

Weight, length and width of the seed and the kernel were determined by taking the average measurement of 20 seeds and kernels. Determinations were done in triplicate. The colour and state of the oil at room temperature were noted by visual inspection.

2.3 Sample Preparation

The fruits were cleaned and decorticated to obtain the seeds. The seeds were dehulled manually, cleaned and separated from the hulls. The kernels were cut into thin slices and then sun-dried at 30 ± 2 °C to constant weight. The dried slices were ground using a mechanical grinder and sieved through a 200 mesh sieve (British standard). The flour obtained was packed in transparent polyethylene bags prior to use.

2.4 Metal Determination

For the metal determination, 0.5 g of each sample was digested with 20 ml of a mixture of concentrated HNO₃ and perchloric acid (2:1 v/v). It decomposes the organic matter completely until the solution becomes a clear one. Thereafter, it was filtered and transferred into a

100 ml standard flask and diluted. It was made up to the mark with deionized water and stored in a clean polyethylene bottle. Sodium and potassium content were determined by using a flame photometer (Model, 405, Corning, UK) as described by Person, (1976), while other mineral element contents were determined using an atomic absorption spectrophotometer (Perkin– Elmer model 703, USA) [19].

2.5 Proximate Analysis

Samples were analysed for dry matter, ash, crude protein, crude fat and total nitrogen according to Association of Official Analytical Chemists methods [20]. Nitrogen content of the seed flour was estimated using the micro-kjeldahl method [21] and the percentage nitrogen was converted to crude protein by multiplying by a factor of 6.25. Carbohydrate contents were determined by difference {100 - (moisture + protein + crude fat + ash + crude fiber)}, [22]. The calorific energy value was obtained by multiplying the values of carbohydrate, protein and crude fat by the Atwater factors of 17, 17 and 37 respectively [23]. Determinations were in triplicate. The crude fat values were used to calculate the total fatty acid by multiplying with a conversion factor of 0.80 [24].

2.6 Feed Compounding

A basal diet was formulated to meet the entire nutrient requirement for young albino rats of 6 weeks (Rattus norvegius). P. longifolia seed flour was used to totally replace wheat flour in the diet formulated (10%). The diets were prepared according to the procedure described by Souza et al. and Ajavi et al. [25-26] with slight modification. The basic ingredients used are as follows: 35% of maize, 18% of soy bean, 3% of calcium, 1% of salt, 15% of groundnut cake, 7.5% of palm kernel cake, 10% of wheat, 7.5% of corn bran and 3% of oyster shell for the control diet). Wheat was totally replaced with 10% of P. longifolia in the experimental diet while 10% of wheat was incorporated in the control diet. Ingredients of the diets were mixed thoroughly with the mixing machine to obtain a homogenous mixture which was pelletized, weighed to be 7000.00 g (100%) for each diet and packed into two different transparent sterile plastic containers for the analysis.

2.7 Animal, Diets and Feeding

Fourteen albino rats (aged 4 - 6 weeks, weighing between 55 g -75 g) were used for this

experiment. They were obtained from the Central Animal House in Veterinary Department. University of Ibadan, Nigeria. The animals were divided into two groups containing seven rats each according to their age gap. They were allowed to acclimatize for one week before the commencement of the experiment. The rats were given feed and water ad libitum for an experimental period of 8 weeks before sacrifice. The experimental rats were fed with a compounded feed where wheat was totally replaced with P. longifolia flour [26]; the control rats were fed with normal feed compounded with wheat flour alone. The physical appearance of both the control and experimental rats was monitored while the body weight of each rat was recorded weekly (without fasting). At the end of feeding period of eight weeks, the rats were fasted overnight and then sacrificed.

2.8 Collection of Sample from Animals

After the eighth weeks of experiment, the rats were sacrificed under mild anesthetics with chloroform after 14-16 h overnight fast. The blood samples were collected by heart puncture into two heparinized tubes for the studies. One tube contains ethylene diamine tetracetic acid with calcium serving as anti-coagulant in the blood sample for haematological analysis while the second tube was stored at -20℃ for the biochemical studies. Serum alanine aminotransferase and aspartate aminotransferase activity were determined by using commercial kits {Randox laboratories Co Atrium, UK}, [27]. Serum creatinine, urea, total globulins protein, albumin, and and albumin/globulin (A/G) were also ratio determined [28]. The internal organs of the rats (liver, kidney, heart, spleen, lungs, intestine, and brain) were collected, weighed and kept in 10% phosphate buffer formalin for histopathology.

2.9 Haematological Analysis

For haematological analysis, 3 ml of blood were collected by cardiac puncture into heparinized tubes and stored at 10°C for analysis the same day. The packed cell volume, haemaglobin concentration, red blood cell and white blood cell counts were determined using standard techniques [29]. The differential WBC counts mean corpuscular volume and mean corpuscular haemaglobin concentration were calculated [30-31]. Microhaemocrit capillary tubes were filled to two-thirds mark with well mixed venous blood. One end was sealed with plasticine. The sealed tubes were placed in microhaematocrit centrifuge and the safety cover securely screwed on. The sealed capillary tubes were centrifuged for 5 mins at 10,000 revolutions per minute. The volume of the red blood cell was read on the micro-haemocrit reader.

2.10 Tissue Pathology

A sample of the liver, kidney, heart and spleen for each animal in the various groups was fixed in 10% phosphate buffer formalin, embedded in paraffin wax, sectioned at 6μ and transferred to clean glass slides. The thin sections were stained with haemotoxylin and eosin (H and E) dyes for examination under the light microscope for histological changes in the tissues due to the consumption of *P. longifolia* and seed flour [30].

2.11 Statistical Analysis

Results were expressed as mean±standard deviation. Organ weights, biochemical and hematological determinations were analyzed by student's t-test. A probability level of p<0.05 was considered significant.

3. RESULTS AND DISCUSSION

3.1 Physical Characteristics

The physical characteristics of *P. longifolia* seeds with regards to the weight, length and width are listed in Table 1. The average weight of 20 seeds is 21.8 ± 0.03 g. The average length and width of the seed are 2.25 ± 0.18 cm and 1.88 ± 0.10 cm respectively. The kernel percentage of 89.90% was obtained for *P. longifolia* seed.

3.2 Chemical Composition

The chemical composition of *P. longifolia* seed flour was viewed in terms of proximate analysis and mineral composition.

The role of trace elements in nutrition cannot be over emphasized. The mineral elements form a small proportion of the total composition of most plant materials and total body weight but do not contribute to the energy values of food. This elements are still of great physiological importance majorly in the body metabolism [32]. The results for the mineral composition in mg/100 g of *P. longifolia* seed flour (Table 1) show that the seeds have high level of iron (706 mg/100 g), followed by manganese (391 mg/100 g); zinc (342 mg/100 g), copper (342 mg/100 g)

and potassium (102 mg/100 g) for P. longifolia. Potassium is an essential mineral element that helps to regulate blood pressure. Calcium, sodium and magnesium were generally low in the seed flours. Nickel, lead and chromium were not detected in the seed flours. The value (0.03) obtained when dividing the concentration of sodium by magnesium for P. longifolia is less than 1. This suggests that the seed flour could probably reduce high blood pressure. At the same time, the value (0.05) obtained for Ca/Mg in P. longifolia seed flour is very low compared with the recommended ratio of 2.2, National Research Council), [33]. This may be caused by the low calcium content of the flour. It may be important to supplement the seed flour with calcium for diet formulation. The results obtained are favorable compared with that of Propolis africana and white melon [34-35].

Table 1. Physical properties and mineral composition of *P. longifolia* seeds

| Physical properties | P. longifolia |
|-----------------------|---------------|
| Weight of 20 seeds(g) | 21.8±0.03 |
| Weight of a seed (g) | 1.09±0.01 |
| Seed length (cm) | 2.25±0.18 |
| Seed width (cm) | 1.88±0.10 |
| Kernel percentage | 89.90% |
| Colour of unripe seed | Green |
| Colour of ripe seed | Black |
| State of the oil @ RT | Liquid |
| Mineral analysis | mg/100 g |
| Sodium | 3.4 |
| Potassium | 102.00 |
| Calcium | 30.30 |
| Magnesiun | 20.90 |
| Manganese | 391 |
| Iron | 706 |
| Copper | 234 |
| Zinc | 342 |
| Nickel | ND |
| Chromium | ND |
| Lead | ND |
| NA/K | 0.03 |
| Ca/Mg | 0.05 |

The chemical composition of *P. longifolia* seed flour was investigated and presented as shown in Table 2. The moisture content and the dry matter of the seed flour are $9.20\pm0.03\%$ and 90.80%respectively. These values obtained are in accordance with the values of 11.00 ± 0.34 and 89.00 obtained for wheat flour [36]. The moisture content obtained is higher than 6.93 for pumpkin, 3.48 in bottle gourd and seed flours [37], 5.00 for white melon seed flour [35] and 1.9 for P. africana [34]. The ash contents obtained from the seed is 2.58±0.02, values which are lower than 3.91 and 4.4±0.1 obtained for Lagenaria siceraria and P. africana [23,34]. The crude protein content of P. longifolia obtained is 12.40±0.25. This value obtained falls within the range of 8.40-14.8 found for cereal seeds such as corn. triticale and wheat [38]. The value is much lower compared with crude protein in rich foods such as 37.1 in Lagenaria seed, [23], 23.6±1.5 in P. africana [34], 27.9 in Parkia biglobos seed [39] and 36.18±0.04 in moringa seed flour [40]. The content of Ρ. longifolia carbohydrate (59.66±0.19) obtained by difference is high. A value of 8.64±0.29 has previously been reported for ginger bread seed flour [41]. P. longifolia seeds might be a good source of carbohydrate and it is an indication that they will be good source of roughage in animal feeds. The ash content is quite low while the moisture is high. The calculated metabolic energy value obtained is 1382.64 KJ/100 g for P. longifolia. This shows that the seed flour could be a source of energy. Table 3 shows the various energy values as contributed by protein, fat and carbohydrate. The proportion of total energy due to protein is 15.26%, proportion due to fat is 11.39% and proportion due to carbohydrate is 73.35% for P. longifolia. This is an indication that the energy obtained from P. longifolia seed is majorly contributed by the high carbohydrate content of the seed flours and could be useful as supplement in compounding animal feed.

3.3 Rat Experiment

3.3.1 Effect of *P. longifolia* seed flour on the proximate composition of the compounded feed

After compounding the feed, diet samples were analyzed again to ascertain their chemical composition. It was then observed that there was an increase in the crude protein, ash and crude fat contents in the diet when compared to the pre-diet compounding proximate values. A decrease was also noticed in the moisture and carbohydrate contents while the dry mater value almost remained the same. Interestingly, moisture, crude protein and crude fat contents were not significantly different in both control and test diets groups (Table 3).

| Parmeters | PLSF ^{\$} | Control group | Test group | Wheat flour [#] |
|-----------------------|-------------------------|-------------------------|-------------------------|--------------------------|
| Moisture content | 9.20±0.03 ^a | 10.15±0.01 ^ª | 8.25±0.01 ^b | 11.00±0.34 |
| Ash content | 2.58±0.02 ^a | 14.1±0.02 ^a | 10.07±0.02 ^b | 1.00±0.04 |
| Crude protein | 12.40±0.25 ^ª | 19.23±0.05 ^a | 19.33±0.12 ^ª | 11.50±0.14 |
| Crude fat | 4.26±0.01 ^b | 15.46±0.01 ^b | 12.5±0.1 [°] | 1.50±0.05 |
| Crude fiber | 11.90±0.04 ^a | 7.58±0.14 ^b | 8.07±0.01 ^a | 0.6±0.01 |
| Carbohydrate | 59.66±0.19 ^ª | 33.46±0.06 ^b | 41.77±0.12 ^ª | 74.90 |
| Dry matter | 90.80 | 89.89 | 91.75 | 89.00 |
| Energy (kj/100 g) | 1382.64 | 1,466.73 | 1,488.11 | 1524.3 |
| Calculated fatty acid | 3.41 | 12.37 | 10.00 | 1.2 |

Table 2. Proximate composition and energy content of *P. longifolia* seed flour, compounded feed and wheat

Mean±SD for three replicate analyses, ^{\$}Values in the same row with the same superscripts are not significantly different at (P< 0.05), [#]Akurbo et al. (2013)

Table 3. Energy values as contributed by protein, fat and carbohydrate of *P. longifolia* seed flour, compounded feed and wheat

| Parameters | PLSF | Control feed | Test feed | Wheat ^a |
|--|---------|--------------|-----------|--------------------|
| Total energy | 1382.64 | 1466.73 | 1488.11 | 1524.3 |
| Proportion of total energy due to protein % | 15.26 | 22.29 | 22.08 | 12.83 |
| Proportion of total energy due to fat % | 11.39 | 39.00 | 31.08 | 3.64 |
| Proportion of total energy due to carbohydrate % | 73.35 | 38.78 | 47.71 | 83.53 |
| $\frac{4}{10}$ Algorithm = -(-1)(0040) | | | | |

[#]Akurbo et al. (2013)

| Table 4. Effect of P. | longifolia seed flour | on rat body w | veight and feed | gain |
|-----------------------|-----------------------|---------------|-----------------|------|
| | | | | |

| Parameters | Control group | Test group |
|----------------------------|---------------|--------------|
| Average initial weight/rat | 55.00±11.9 | 55.00±5.00 |
| Average final weight/rat | 141.58±16.18 | 145.00±17.08 |
| Weight gain/rat | 86.58 | 90.00 |
| % weight gain | 157.42 | 163.63 |
| Survival rate (s) % | 100 | 100 |

3.3.2 Physical appearance of test and control rats

Weekly inspection of the physical appearance of the rats revealed that the rats were normal in all the groups throughout the eight weeks of the experiment. The eyes, mouth and hair of the animals in both groups appeared to be normal throughout the period of the study. No mortality was recorded for any of the groups.

3.3.3 Body weight and weekly feed consumption of the rats

Figs. 1 and 2 show the weekly feed consumed and the body weight of the different groups of rats. Fig. 1 revealed that the feed consumed increased gradually along the weeks of the experiments. The total feed consumed by the rats in the tests groups are a little higher than those consumed by the rats in the control groups. In Fig. 2, all the test rats steadily gained weight during the period of the experiment. The average weight gain per rat is shown to be 86.58 g for control and 90.00 g for test group respectively. This indicated that the rats in the test group had a higher average weight gain than those in the control groups.

3.3.4 Organ weights

The organs whose weights were taken are brain, liver, heart, kidney, lungs, spleen and intestine (Fig. 3). At the moment of sacrifice, organs of the test and control rats which are similar have comparable weights; the seed flour has no significant effect (p<0.05) on the weight of similar organs of the test and control group of rats. Liver weight was 4.74 ± 0.63 g for the control and 4.62 ± 0.63 g for test. A value of 3.92 ± 0.19 g to 3.95 ± 0.21 g has been previously reported as liver weights for rats fed with crude

Balanite aegyptiaca seed oil in rat [42]. The liver weight of 6.84 ± 1.08 g (control group) to 6.17 ± 1.15 test group) was reported for rat fed with Greenwayodendron suavelens seeds [43].

3.4 Haematology, Blood Chemistry and Histopathology

3.4.1 Haematological analysis

There was no significant difference ($p \le 0.05$) in the result of the haematological analysis of test and control groups (Table 5) of rat fed with *P. longifolia* seed flour. The rats were not anaemic, their PCV values are similar to those reported for healthy murine species [44]. Therefore, the diet compounded with *P. longifolia* seed flour might have no adverse effect on the blood of the rats under this study as it is comparable to the indices obtained for the diet compounded with wheat. This was in accordance with previous result [45-46]. The similarity and the closeness of the WBC counts observed in the control groups when compared to the corresponding test groups suggests that the rats had no infection. This is similar to previous reported work on *T. occidentalis* [47]. The platelets (platelets cells/cu.mm) x 10⁶) values obtained were 0.150±0.05 and 0.115±0.03 for control and test groups respectively while the haemoglobin



Fig. 1. A chart showing the weekly feed consumption of rats fed with *P. longifolia* seed flour feeds. Series 1 is the control while series 2 is the test group



Fig. 2. A chart showing the weekly body weight (g) of rats fed with *P. longifolia* seed flour feed. Series 1 is the control while series 2 is the test group



Fig. 3. A chart showing the mean internal organ weight (g) of rats fed with *P. longifolia* seed flour feed. Series 1 is the control while series 2 is the test group. Brain, liver, kidney, heart, lungs, spleen and intestine are represented respectively by 1, 2, 3, 4, 5, 6 and 7 along the y axis

values are 16.73 ± 1.05 and 15.42 ± 1.13 respectively for control and test groups. The haematological values from this study are similar to the result reported on the toxicity study of *Garcinia mangostana* pericarp extract in rats [45].



Fig. 4. Photomicrograph of the kidney of test group rats fed with *P. longifolia* seed flour showing no visible lesion. (x550)

3.4.2 Blood chemistry

Table 5 also revealed the result of blood biochemistry of the rats fed with *P. longifolia* seed flour. The serum protein values are comparable to each other in the corresponding control and test groups. Alanine aminotransferases enzyme activity examined

was 28.33 ± 0.94 and 30.00 ± 2.82 respectively for control and test. Alanine aminotransferases enzyme activity value of 20.33 ± 9.87 in the case of the experimental feed as against 15.14 ± 6.41 for the control was previously reported for *Greenwayodendron suavelens* seeds in rat feed [43].



Fig. 5. Photomicrograph of the kidney of control group rats fed with feed without *P. longifolia* seed flour showing no lesion (x550)

3.4.3 Histopathological analysis

No lesion was observed in the heart, kidney and liver of the rats in the control and test groups (Table 6). This is an indication that *P. longifolia* seed flour was probably not harmful to the organs of the rats.

| Haematological analysis* | Control group | Test group |
|---|--------------------------|-------------------------|
| PVC (%) | 46.86±2.54 ^a | 46.57±2.82 ^a |
| MCHC (%) | 33.11±0.66 ^ª | 34.22±0.36 ^a |
| MCH (%) | 20.1±0.53 ^a | 23.42±0.05 ^a |
| MCV (%) | 60.72±1.48 ^a | 62.21±8.33 ^a |
| Hb (mg/de) | 16.73±1.05 ^a | 15.42±1.13 ^ª |
| RBC (10 ⁶ /ml) | 8.14±0.60 ^a | 7.68±0.62 ^a |
| WBC (10 ³ /ml) | 8,793±0.79 ^a | 7,607±2.9 ^a |
| Lymplocyte (%) | 79.001±9.35 ^a | 72.00±5.38 ^a |
| Neutrophil (%) | 20.57±8.83 ^a | 23.14±3.74 ^ª |
| Monocyte (%) | 2.43±1.27 ^a | 2.57±1.72 ^a |
| Eosinophil (%) | 2.00 ±1.15 ^ª | 2.86±3.83 ^a |
| Platelets (cells/cu.mm)x10 ⁶ | 0.150±0.05 ^ª | 0.115±0.03 ^a |
| Biochemical analysis* | | |
| TP (g/dl) | 7.86±0.30 ^a | 6.77±0.70 ^a |
| ALB (g/dl) | 5.07±0.22 ^a | 4.07±0.60 ^a |
| GLB (g/dl) | 2.33±0.60 ^a | 2.67±0.11 ^a |
| AL/GLB | 2.17±0.57 ^a | 1.52±0.2 ^b |
| AST (g/l) | 40.00±1.00 ^a | 41.33±2.43 ^a |
| ALT (g/l) | 28.33±0.94 ^a | 30.00±2.82 ^a |
| ALP (g/l) | 80.33±6.67 ^a | 73.33±1.15 ^ª |
| Urea | 14.67±0.71 ^a | 15.00±1.00 ^a |
| Creatinine | 0.73±0.1 ^a | 0.6±0.10 ^a |

 Table 5. Haematological and biochemical studies of rats fed with control feed and

 P. longifolia seed flour

Hb = Haemoglobin, concerntration (g%); PCV = Packed cell volume (%), RBC = Red Blood Cell Counts,
 WBC = White Blood cell count (x10³/mm3), MCV = Mean Corpuscular Volume (fl), MCH = Mean Corpuscular Haemoglobin (%); MCHC = Mean Corpuscular Haemoglobin Concentration (%). AST- Aspartate aminotransferases, ALT- Alanine aminotransferases, ALP = Alkaline phosphatase; ALB = Albumin; GLB = Globulin; ALB/GLB = Albumin – Globulin ratio; TP = Total Protein

*Values in the same row with the same superscripts are not significantly different at (P< 0.05)

Table 6. Histological result of tissues from the rats fed with *P. longifolia* seed flour

| Tissue | Control group | Test group |
|--------|-------------------|-------------------|
| Kidney | No visible lesion | No visible lesion |
| Liver | No visible lesion | No visible lesion |
| | seen | seen |
| Heart | No visible lesion | No visible lesion |
| | | |

4. CONCLUSION

P. longifolia seed flour could be utilized successful as source of dietary supplement in feed for livestock due to the protein content and the high carbohydrate values. Their low protein content can still be supplemented with other high protein residue such as groundnut cake, soy bean cake, *B. aegyptiaca* cake and melon cake. *P. longifolia* seed flour is very high in iron,

manganese, zinc, copper and potassium but low in calcium, sodium and magnesium. Therefore, it might be important to supplement the seed flour with calcium for diet formulation.

P. longifolia seed flour, when fed to rat, did not have any deleterious effect on the physical appearance, body weight gain, organ weights, haematological and biochemical parameters. The seed flour was found not to be toxic to the heart and kidney of the rats. No lesion was seen in the liver of the rat fed with *P. longifolia*. It is then suggested that the seed flour might be a good source of nutrient for livestock diet formulation. *P. longifolia* seed flour can still be studied at graded level of incorporation in order to determine the suitable level or percentage at which the seed flour could possibly and effectively be integrated in formulating feed for livestock.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Singh B, Singh U. Peanut as a source of protein for human foods. Plant Foods for Human Nutrition. 1991;41:165-177.
- Giami SY, Wachuku OC. Composition and functional properties of unprocessed and locally processed seeds from three underutilized food sources in Nigeria. Plant Food for Human Nutrition. 1997;50:27-36.
- Enujiugha VN. Development of a new food paste from seeds of *Pentaclethra* species. Applied Tropical Agriculture. 2000;5:89-94.
- Enujiugha VN, Ayodele-Oni O. Evaluation of nutrients and some anti-nutrients in lesser known, underutilized oilseeds. International Journal of. Food Science and Technology. 2003;38:525-528.
- Eddy NO, Udo CL. The energy value of some Nigerian soup. Pakistan Journal of Nutrition. 2004;3:101-103.
- Amaechi NC. Nutritive and anti-nutritive evaluation of wonderful kola (*Buccholzia coricea*) seeds. Pakistan Journal of Nutrition. 2009;8:1120-1122.
- Onwereuzo JC, Obanu ZA, Onuoha KC. Functional properties of some lesser known tropical legumes. Journal of Food Science and Technology. 1994;31:302-306.
- Kar S, Maitya JP, Samal AC, Santra SC, Jean JS. Bundschuh journal of deposition and uptake of metals in urban canopy: Atmospheric arsenic sequestration. Journal Hazard Mat; 2013. Available:<u>http://dx.doi.org/10.1016/j.jhazm</u> at.2012.12.048
- Rastogi RP, Mehrotra BN. Compendium of Indian Medicinal Plants Central Drug Research Institute Lucknow & NISC, New Delhi, CSIR. 1998;5.
- Saleem R, Ahmed M, Ahmed SI, Azeem M, Khan RA, Rasool N. Hypotensive activity and toxicology of constituents from root bark of *Polyalthia longifolia*. Phytotherapy Resource. 2005;19:881-884.
- 11. Nair R, Shukla V, Chanda S. Assessment of *Polyalthia longifolia* var. pendula for hypoglycemic and anti-hyperglycemic activity. Journal of Clinical Diagnosis Resources. 2007;3:116-121.

- Annapuma Y, Shaktimitra DA, Iyengar S, Rao N, Rao UTB. Antimicrobial activity of leaf extracts of *Polyalthia longifolia*. Journal of Phytopathology. 1983;106:183-185.
- 13. Faizi S, Khan RA, Azher S, Khan SA, Tauseef S, Ahmad A. New antimicrobial alkaloids from the roots of *Polyalthia longifolia* var. pendula. Planta Medicinal Journal. 2003;69:350-355.
- Murthy MM, Subramanyam M, Bindu MH, Annapurna J. Antimicrobial activity of clerodane diterpenoids from *Polyalthia longifolia* seeds. Fitoterapia. 2005;76:336-339.
- Ghosh A, Bidus KD, Soroj KC, Goutam C. Antibacterial potentiality and hytochemical analysis of mature leaves of *Polyalthia longifolia* (*Magnoliales: Annonaceae*). The South Pacific Journal of Natural Science. 2008;26:62-67.
- FAO, FAO/WHO/UNU. Energy and protein requirements: Report of a Joint FAO/WHO/UNU Expert Consultation; 1985.
- 17. Gustafsson E, Edlund M, Hagberg M. Effect of food additives. Health Beliefs. 2003;34:565-570.
- Oyedeji FO, Adeleke BB, Akintola CB. Physicochemical and fatty acid profile analysis of *Polyalthia longifolia* seed oil. Trends in Applied Science Resources. 2011;6:614-621.
- Onyeike EN, Acheru GN. Chemical composition of selected Nigerian oil seeds and physicochemical properties of the oil extracts. Journal of Food Chemistry. 2002; 77:431-437.
- AOAC. Official method of analysis of AOAC. 17th edition. Association of Official of Analytical Chemist International, Viginia, U.S.A. 2005;1-37.
- Pearson D. The Chemical Analysis of Foods 7th edition Churchill Livingstone. 1976;488-496.
- Ajayi IA, Dawodu FA, Adebowale KO, Oderinde RA. Chemical composition of *Pentaclethra macrophylla* seed and seed oil grown in Nigeria. La Rivista Italiana Delle Sostanze Grasse. 2002;76:183-185.
- 23. Olaofe O, Ekuagbere AO, Ogunlade L. Chemical, amino acid composition and functional properties of calabash seed's kernel. Bulletin of Pure and Applied Science. 2009;28:13-24.

- Grennfeild H, Southgate D. Food composition data production, management and uses, 2nd edition, F A O, Rome, 223; 2003.
- Souza ARde, Martins LP, Faira LCde, Martins MEP, Fereira RN, Silva AMLda, Gil ES, Conceição ECda. Studies on the bioavailability of zinc in rats supplemented with two different zinc-methionie compounds. Latin American Journal of Pharmacy. 2007;26:825-830.
- 26. Ajayi IA, Ifedi E, Vivian NA. Amino acid analysis and preliminary toxicological evaluation of *Garcinia mangostana* seed cake. Global Sciences Journal. 2013; 13:17-21.
- Chawla R. Practical clinical biochemistry (Methods and Interpretations). Second edition. Jaypee Brothers Medical Publishers, New Delhi, India. 1999;106-118.
- Dacie JV, Lewis SM. Practical, 7th Edition. Churchill Livingstone, Edinburg, London. 1995;12-17.
- 29. Jain NL. Schalmes veterinary haematology. 4th Edition, Lea and Ferbiger, Philadelphia. 1986;3:281.
- Ajayi IA, Oderinde RA, Ogunkoya BO, Egunyomi A, Taiwo VO. Chemical analysis and preliminary toxicological evaluation of *Garcinia mangostana* seeds and seeds oil. Journal of Food Chemistry. 2007;101:999-1004.
- 31. Schwart MK. Role of trace elements in cancer. Cancer Research. 1975;35:3481-3484.
- National Research Council Recommended Dietary Allowances, 10th edition; 1989. National academic Press, Washington, D. C. USA.
- Aremu MO, Olonisakin A, Atolaye BO, Ogbu CF. Some nutritional composition and functional properties of *Prosopis africana*. Bangladesh Journal of Science Industrial Research. 2007;42:269-280.
- Eunice MO, Fagbemi NT, Osundahunsi FO. Chemical and functional properties of full and defatted white melon (*Cucumeropsis mannii*) seed flours. Journal of Food Science and Engineering. 2012;2:691-698.
- 35. Akubor PI, Yusuf D, Obiegunam JE. Proximate composition and some functional properties of flour from the

kernel of African star apple (*Chrysophyllum albidum*). International Journal of Agricultural Policy and Research. 2013;1:062-066.

- Olaofe O, Adeyemi FO, Adediran GO. Amino acid and mineral compositions and functional properties of some oilseeds. Journal of Agricultural and Food Chemistry. 1994;42:878-881.
- Heger J, Eggum BO. The nutritional values of some high yielding cultivars of tricale. Journal of Cereal Science. 1992;14:63-71.
- Elemo GN, Elemo BO, Oladunmoye OO, Erukainure OL. Comprehensive investigation into the nutritional composition of dehulled and defatted African locust bean seed (*Parkia biglobosa*). African Journal of Plant Science. 2011;5:291-295.
- Singh B, Singh U. Peanut as a source of protein for human foods. Plant Foods for Human Nutrition. 1991;41:165-177.
- Tidjani A, Issoufou A, Mohamed T, Kamara KZ, Huiming Z. Chemical and nutrient analysis of gingerbread plum (*Neocarya macrophylla*) seeds. Advance Journal of Food Science and Technology. 2010;2:191-195.
- 41. Obidah W, Nadro MS, Tiyafo GO, Wurochkke AU. Toxicity of crude *Balanite aegyptiaca* seed oil in rats. Journal of American Science. 2009;5:13-16.
- 42. Ajayi IA, Olaifa FE, Omoniyi MM. Chemical analysis and nutritional assessment of defatted *Garcinia mangostana* seeds used as an additive on the feed of fish (*Clarias gariepinus*). Global Journal of Science Frontier Research Chemist. 2013;13:38-45.
- Oyewale JO, Olayemi FO, Oke OA. Haematology of the wild adult African giant rat (*Cricetomys gambianus* water house). Veterinary Archive. 1988;68:91-98.
- 44. Vishnu PV, Sankari G, Mallika J, Surapaneni KM, Aishwarya TS, Saraswathi P, Chandra S, Gopan VS. Auto fluorescence and Fourier transform – infra red spectral investigation on diethyl nitrosamine (Den) induced hepatocellular carcinoma, treated with pericarp extract of *Garcinia mangostana* Linn in rats. Journal of Clinical and Diagnostic Research. 2010; 4:3289-3297.
- 45. Ajayi IA, Aghanu VN, Antia RW, Marchini SJ. Evaluation of *Monodora tenuifolia* seed

Ajayi and Ifedi; ACSJ, 15(3): 1-12, 2016; Article no.ACSJ.19789

oil. Annals of Food Science and Technology. 2012;13:61-65.

46. Ajayi IA, Oderinde RA, Taiwo VO, Agbedana EE. Dietary effect on growth

plasma lipid and tissues of rats fed with non-conventional oil of *Telfairia occidentalis*. Journal of Food Science and Agriculture. 2004;84:1715-1721.

© 2016 Ajayi and Ifedi; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: http://sciencedomain.org/review-history/15303