



## **Chemical Composition of *Pinus brutia* Ten Essential Oil and Its *in vitro* Anti-Inflammatory Activity**

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### **Authors' contributions**

*This work was carried out in collaboration among all authors. Author LK performed the experiments; wrote the paper; analyzed and interpreted the data. Author RN designed the experiments; analyzed and interpreted the data. All authors read and approved the final manuscript.*

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### **ABSTRACT**

This investigation aims to determine the chemical composition of *Pinus brutia* leaves essential oil and evaluate its anti-inflammatory property using Human Red Blood Cells (HRBC) membrane stabilization assay and Albumin denaturation assay. The chemical composition of essential oil (EO) obtained by hydro-distillation of leaves of *Pinus brutia* was investigated by GC-MS. The anti-inflammatory effect of EO was evaluated using Human Red Blood Cells (HRBC) membrane stabilization assay and Albumin denaturation assay. The main constituents of EO were  $\alpha$ -Terpineol (66.16%), 3-Carene (4.90%), Carveol (4.55%) and cis-Verbenol (3.22%). The inhibition of hemolysis was observed at concentrations (2.5-12.5)  $\mu$ g/ml. Moreover, albumin denaturation test showed protection effect at concentrations (8-40)  $\mu$ g/ml. We concluded that, *Pinusbrutia* EO shows strong anti-inflammatory activity at different concentration when compared to standard drug of Diclofenac sodium. In addition, GC-MS analysis of *Pinus brutia* EO showed the presence of  $\alpha$ -Terpineol as major compound in the oil. It reveals that this constituent is responsible to maximum protection of albumin denaturation and membrane stabilization assay. The future work will be determination of anti-inflammatory by *in vivo* models.

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## 1. INTRODUCTION

Essential oils, as defined by the European Pharmacopeia 7th edition, are "Odorant products, which have the complex composition, and obtained from plant raw extract, either extracted by steam of water, dry distillation or a suitable mechanical method without heating. Generally, a physical method is used for the separation of essential oil from the aqueous phase which has no significant change in its chemical composition" [1].

Essential oils found in many different plants, especially the aromatic and characterized in a number of families such as: Asteraceae, Lamiaceae, Lauraceae, Myrtaceae, Rutaceae, Cupressaceae and Piperaceae [2]. However, parts of plants, which serve as the major source of essential oil can be different. Those include roots (valerian), peels (lemon), leaves (mint, pine), fruits (black pepper), barks (Cinnamon) and flowers (chamomile) [3].

Essential oils compounds can be divided into two main groups: terpene hydrocarbons (monoterpenes and sesquiterpenes) and their oxygenated compounds such as: Phenols, Aldehydes, Ketones, Esters, Lactones and Ethers [4].

Many methods can be used to extract essential oils, which are dependent on botanical material used. Extraction techniques can be divided into two categories:

1. Classical methods (Steam distillation, Hydro-distillation and Solvent extraction).
2. Innovative methods Such as supercritical fluid extraction and Microwaves assisted extraction [5].

Essential oils have many applications in various industries, such as food products, drinks, perfumes, pharmaceuticals and cosmetic [6]. Therapeutic properties varies from plants to another, in Table 1 some examples of essential oils and their main active compounds as well as their therapeutic effects:

*Pinus brutia* Ten. (Pinaceae) is known by several other names, Turkish pine, Calabrian pine (from a naturalized population of the pine in Calabria, Southern Italy, from where the pine was first botanically described), East Mediterranean pine and Brutia pine. *Pinus* trees are used in many industries; the turpentine which is obtained mainly from pine trees is used in medicine, pharmacy, food, cosmetics, paint and coatings [36].

Moreover, the resin obtained from *Pinus brutia* was used traditionally to treat Stomach ulcer, cough and it was also applied externally to heal wounds [37].

According to ethnobotanical study of *Pinus* species in Turkey, 130 traditional medicinal and ethnobotanical studies published up to 2011 which were dealt different areas of Turkey are examined and the usages of *Pinus* species are compiled from 54 of them. It has 269 records that are to be proof of the wide range of ethnobotanical usages of pines. It is stated in the literature that the most important medicinal usage of *Pinus* species is for respiratory system diseases and inflammatory diseases [37].

Nature-based medicines are having increased attention in the quest for novel pharmacophores that hold the prospect of enhanced therapy.

This interest follows the World Health Organization's 2008 ratification of The Beijing Declaration, which promotes the safe and effective use of traditional and alternative medicines and claims greater assimilation of these into national health care systems in order to improve therapy with less cost and side effects [38].

Therefore, this study aims to evaluate the anti-inflammatory activity and determine the active compounds which may be responsible for this activity.

In this study, the essential oil composition of the leaves of *P. brutia* collected from Syria were analyzed by GC/MS system, and its anti-inflammatory property was tested in-vitro using HRBC test and Albumin denaturation assay.

**Table 1. Examples of essential oils and their main active compounds as well as their therapeutic effects**

No	Essential oil	Main active compounds	Properties
	Chamomille essential oil	Bisabolol and chamazulene [7,8]	Anti-inflammatory, anti-allergic, decongestive (decongest the skin), anti-pruritic healing and antispasmodic [9]
	Dill essential oil	Carvone. [10]	Antispasmodic in gastrointestinal disorders [11]
	Garlic essential oil	Diallylle disulfide [12,13]	Maintains and protects the cardiovascular system, hypoglycemic, Regulates blood pressure vermifuge, antimicrobial, antiviral, anti-fungal and anti-parasitic, insecticidal and larvicidal, antioxidant [14]
	Clove essential oil	Eugenol and eugenyle acetate [15]	Antimicrobial, antiviral, antifungal, general stimulating, hypertensive aphrodisiac, carminative, anesthetic [16–19]
	Cinnamon essential oil	Cinnamaldehyde [20]	Antibacterial, antiviral, antifungal and parasiticide [21]
	Eucalyptus essential oil	1, 8-cineole [22]	Anticatarrhale, mucolytic and expectorant, antimicrobial, Antiviral [23–25]
	Peppermint essential oil	Menthol and menthone [26]	Tonic and stimulant, decongestant, anesthetic and analgesic antipruritic, refreshing, antimicrobial, anti-inflammatory, expectorant, mucolytic, emmenagogue [27–30]
	Lavender essential oil	Linalol and linalyle acetate [31,32]	Antispasmodic, relaxing, sedative, analgesic, anti-inflammatory and antimicrobial [33]
	Tea tree essential oil	Terpinene-1-ol-4 [34]	Antimicrobial, antiviral, antiasthenic, neurotonic, lymphatic, decongestant, radioprotective, antispasmodic [35]

## 2. MATERIALS AND METHODS

### 2.1 Instrumentation and Apparatus

1	Electronic balance (Sartorius AG, Germany)
2	Ultrapure TM water purification system (Lotun Co., Ltd., Taipei, Taiwan)
3	UV-1800 spectrophotometer (Shimadzu, Japan)
4	Water bath
5	Gas chromatography/mass spectrometry(Agilent, United States)

### 2.2 Materials and Reagents

**Chemical materials:** Sodium phosphate dibasic dehydrate (sigma Aldrich, Germany), Sodium phosphate monobasic dihydrate (Acros organics, United States), Distilled deionized water, Dimethyl sulfoxide- DMSO (sigma Aldrich, Germany), Sodium Chloride (HiMedia Laboratories, India) and Sodium Diclofenac (Amoli Organics Pvt. India).

**Plant source:** The leaves of *Pinus brutia* were collected in March from classified trees growing in the campus of Aleppo University. The leaves were dried in the shade in a well-ventilated place, then stored in airtight containers.

### 2.3 Methods

#### 2.3.1 Isolation of the essential oil

Air-dried leaves (100 g) were subjected to hydro distillation using a Clevenger-type apparatus for 3h.

#### 2.3.2 Gas chromatography and mass spectrometry (GC-MS) analysis

The oil was analyzed by GC-MS, using a Hewlett Packard system. HP-Agilent 5975 N GC-MS system with 7890 GC in Aleppo University.

Method was done according to Bagci et al with some modification: [39]

DB-5 HT column (30 m × 0.32 mm i.d., film thickness 0.1µm) was used with helium as the carrier gas. Injector volume was 10µl, split ratio was 1:10, and front inlet temperature was 290 °C. The GC oven temperature was kept at 60°C for 0 min and programmed to 280°C at a rate of

5°C/min and then kept constant at 280°C for 1 min.

Component identification was carried out using spectrometric electronic libraries (WILEY, NIST).

#### 2.3.3 Evaluation of anti-inflammatory activity

The anti-inflammatory activity of essential oil of *Pinus brutia* leaves was evaluated by human red blood cell (HRBC) membrane stabilization and albumin denaturation assay.

##### 2.3.3.1 HRBC membrane stabilization assay

The effects of the essential oil on hemolysis of HRBC induced by heat was evaluated using the method of Shinde et al. with some modifications [40].

##### 2.3.3.1.1 Preparation of erythrocyte suspension

Fresh whole blood (3 ml) was collected from healthy volunteers, who were nonsmokers, did not take alcoholic drinks, and did not use any chemical medicine for one week; the samples were put into heparinized tubes then centrifuged at 3000 rpm for 10 min. A volume of normal saline equivalent to that of the supernatant was used to dissolve the red blood cells. The volume of the dissolved red blood cells obtained was measured and reconstituted as a 40% suspension with isotonic buffer solution (10 mM sodium phosphate buffer, pH 7.4). The buffer solution contained 0.2g of NaH<sub>2</sub>PO<sub>4</sub>, 1.15 g of Na<sub>2</sub>HPO<sub>4</sub> and 9 g of NaCl in 1liter of distilled water.

##### 2.3.3.1.2 Assay of membrane stabilization by heat induced hemolysis

Essential oil was dissolved by Dimethyl sulfoxide (DMSO) 10% to obtain concentration (5-10-15-20-25 µg/ml).

Two groups of centrifuge tubes were prepared in such a way that each tube contained 5 mL of essential oil, 4.85 mL of isotonic buffer solution and 0.15mL of HRBC suspension 40%. One of the group was incubated in a water bath at 54°C for 20 minutes. The other group was placed in the refrigerator. The negative control was prepared by putting saline instead of essential oil and Sodium Diclofenac was used as a standard drug.

Afterwards, all tubes were centrifuged at 3500 rpm for 7 min and the absorbance of the supernatants was taken at 560nm. The experiment was performed in triplicate. Percentage of membrane stabilization activity was calculated by the formula mentioned below:

$$\text{Protection \%} = 1 - \left( \frac{\text{OD}_2 - \text{OD}_1}{\text{OD}_3 - \text{OD}_1} \right) * 100$$

Where:

OD1= absorbance of test sample unheated  
 OD2 = absorbance of test sample heated  
 OD3=absorbance of control sample heated.

#### 2.3.3.2 Evaluation of essential oil effect on albumin denaturation

The effects of the *Pinus brutia* essential oil on Albumin denaturation was evaluated using the method of Chatterjee et al. with some modifications [41].

The reaction mixture contains 2.8 ml of isotonic phosphate buffer (10 mM sodium phosphate buffer, pH 7.4), 0.2 ml of egg albumin (from hen eggs) and 2 ml of different concentrations from Essential oil (20-40-50-60-100) µg/ml. Distilled water was used as a negative control and Sodium Diclofenac was used as a positive control.

The reaction mixture was incubated at 37°C for 20 minutes then incubated at 70°C for 5 minutes. After cooling, the absorbance was measured at 660nm, the percentage of protection of protein denaturation was calculated as in the following equation: [42]

$$\text{Protection\%} = 1 - \left( \frac{\text{studied sample absorbance}}{\text{negative control absorbance}} \right) * 100.$$

#### 2.3.4 Statistical study

All experiments were performed triplicates; results were expressed as mean values ± standard deviation (SD). Statistical analysis was carried out using the Statistical Package for the Social Science SPSS Version 22. Results were statistically analyzed by one-way ANOVA. The p-value<0.05 was statistically significant when compared with control.

### 3. RESULTS AND DISCUSSION

#### 3.1 GC-MS Analysis

Yield of oil was 0.38%, Gas chromatogram of the essential oil from dried leaves is shown in Fig. 1. The identified constituents of the essential oils are listed in Table 2.

In the essential oil of *Pinus brutia* leaves, the main components were identified and constituting 82.93%.

The major components of *Pinus brutia* leaves were α-Terpineol (66.16%), 3-Carene (4.90%), Carveol (4.55%) and cis-Verbenol (3.22%).

Experimental results from our study, concerning the composition of essential oils, are in accordance with previously published data. The composition of the essential oils isolated from the flowers and cones of *Pinus brutia* grown in Lebanon were investigated and the main components were monoterpenes and oxygenated monoterpenes such as α-Pinene, β-Pinene and Terpinen-4-ol [43].

On the other hand, according to a study which was carried out in Tunisia, the essential oil of *Pinus brutia* leaves was characterized with presence of further component such as: Thujene and phellandrene [44].

#### 3.2 Assay of Membrane Stabilization by Heat Induced Hemolysis

Essential oil of *Pinus brutia* Leaves showed efficiency in membrane stabilization by heat induced hemolysis as shown in Table 3. Whereas the oil showed the highest efficiency which reached 56.71% at concentration 10µg/ml.

Lysosomes are one of the factors that may contribute to tissue damage during the inflammatory process, by oxidizing cell membrane lipids.

In 1979, Studies also showed that lysosomes inhibit the steroid anti-inflammatory receptor Hsp90 by changing it to a smaller form, preventing its joining with the steroidal anti-inflammatory drug, which increases the inflammatory state [45].

Therefore, the stabilization of the lysosome membrane contributes in preventing the mediators release such as proteases and

reducing the inflammatory response. The erythrocyte membrane is similar to the lysosome membrane. Thus, the anti-hemolytic effect of plant essential oil can be taken as evidence of the anti-inflammatory effectiveness [42].

### 3.3 Evaluation of Essential Oil Effect on Albumin Denaturation

Essential oil showed efficiency in albumin denaturation assay as shown in Table 4. The oil showed the highest efficiency which reached 55.38% at concentration 40 µg/ml, while sodium Diclofenac did not give a noticeable efficacy at the concentration of 40 µg/ml.

Researchers found that denaturation of protein is one of the causes of rheumatoid arthritis. Production of auto-antigen in certain arthritic

diseases may be due to denaturation of protein [46].

The anti-inflammatory property of essential oils may be explained by considering that essential oil are able to scavenge some free radicals, which play major role in inflammatory response [47] and this is in accordance with traditional use where *Pinus* species have been used against rheumatic pain and inflammatory cases [48].

Furthermore, in a study carried out to evaluate the anti-inflammatory property of alpha-terpineol, revealed that alpha-terpineol had an inhibiting effect on IL-6 formation. This anti-inflammatory effect of alpha-terpineol on IL-6 formation was verified by quantitative real-time reverse transcription Polymerase Chain Reaction experiments in which alpha-terpineol inhibited the gene expression of the IL-6 receptor [49].

**Table 2. Percentage composition of essential oil from *Pinus brutia* leaves**

No	Compound	Percentage%
	α-Terpineol	66.16
	Carveol	4.55
	Berbenol	1.05
	Aromadendrene	2.10
	cis-Verbenol	3.22
	3-Carene	4.90
	Cinen	0.95

**Table 3. Effect of *Pinus brutia* essential oil on heat induced hemolysis of HRBCs**

Treatment	Concentration µg/ml	Protection% Mean±SD
<i>Pinus brutia</i> Essential oil	2.5	48.04±8.36
	5	49.03±1.82
	7.5	55.80±1.39
	10	56.71±2.98
	12.5	55.71±2.73
Sodium Diclofenac	100	89.21±2.54

**Table 4. Effect of *Pinus brutia* essential oil on albumin denaturation**

Treatment	Concentration µg/ml	Protection% Mean±SD
<i>Pinus brutia</i> Essential oil	8	15.25±3.05
	16	22.7±6.11
	20	24.17±1.27
	24	31.51±2.06
	40	55.38±4.16
Sodium Diclofenac	40	-
	120	11.02±2.70
	160	22.80±1.30

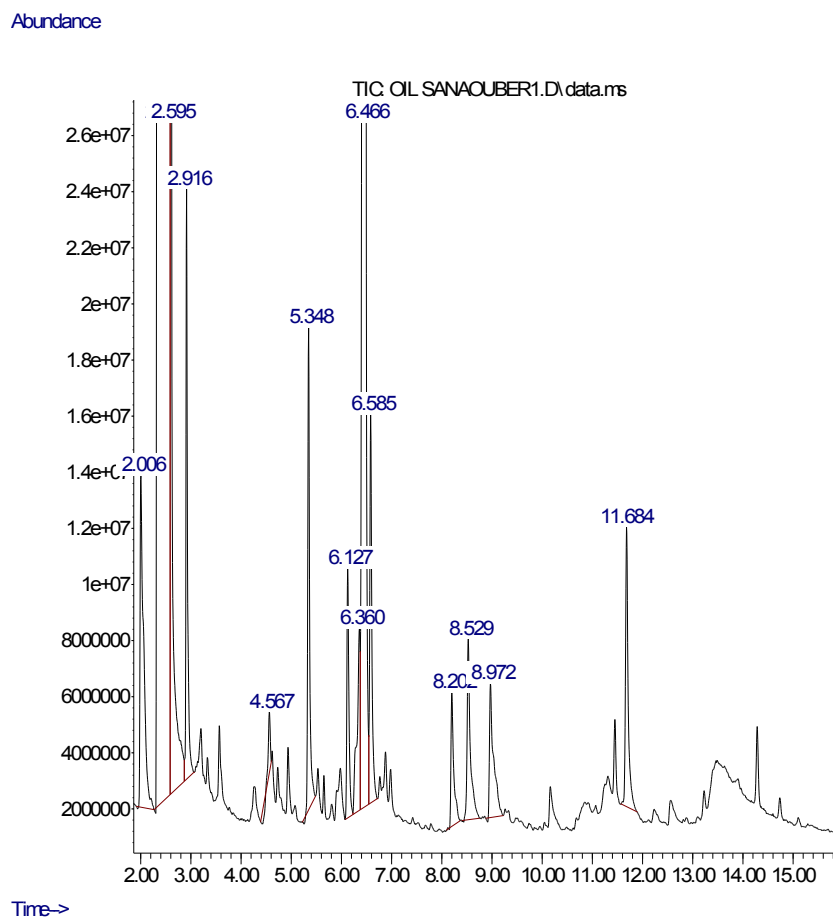


Fig. 1. Gas chromatogram of the essential oil from dried leaves of *Pinus brutia*

#### 4. CONCLUSION

The GC/MS analysis proves that the major compound of *Pinus brutia* leaves essential oil was  $\alpha$ -Terpineol, which may attribute to the anti-inflammatory activity of EO. The in-vitro anti-inflammatory tests which were carried out in this study indicate strong anti-inflammatory effect of *Pinus brutia* EO: further in vivo studies are needed to ensure this activity.

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#### COMPETING INTERESTS

Authors have declared that no competing interests exist.

#### REFERENCES

1. Council of Europe; European pharmacopoeia commission; European directorate for the, quality of medicines & healthcare. European Pharmacopoeia. 7th ed. Strasbourg: Council Of Europe: European Directorate for the Quality of Medicines and Healthcare; 2010.
2. Joy PP. Tropical aromatic and medicinal plants; 1998.
3. Tongnuanchan P, Benjakul S. Essential oils: Extraction, bioactivities and their uses for food preservation. J Food Sci. 2014;79(7):R1231-R1249. DOI: <https://doi.org/10.1111/1750-3841.12492>
4. Aziz Z, Ahmad A, Mohd-Setapar S, et al. Essential oils: Extraction techniques, pharmaceutical and therapeutic potential - A review. Curr Drug Metab. 2018;19.

- DOI:10.2174/1389200219666180723144850
5. Mejri J, Abdelkarim A, Mejri M, Abderrabba M. Emerging extraction processes of essential oils: A review. *Asian J Green Chem.* 2018;2.  
DOI:10.22631/AJGC.2018.119980.1053
  6. Jilani A, Dicko A. The therapeutic benefits of essential oils. In: *Nutrition, well-being and health*; 2012.  
DOI: 10.5772/25344
  7. Cemek M, Kağa S, Şimşek N, Büyükkuroğlu M, Konuk M. Antihyperglycemic and antioxidative potential of *Matricaria chamomilla* L. in streptozotocin-induced diabetic rats. *J Nat Med.* 2008;62:284-293.
  8. Kamatou GPP, Viljoen AM. A review of the application and pharmacological properties of  $\alpha$ -bisabolol and  $\alpha$ -bisabolol-rich oils. *J Am Oil Chem Soc.* 2010;87(1):1-7.  
DOI:10.1007/s11746-009-1483-3
  9. Bnouham M. Medicinal plants with potential galactagogue activity used in the moroccan pharmacopoeia; 2010.  
DOI:10.2202/1553-3840.1268
  10. Genotoxicity of dill (*Anethum graveolens* L.), peppermint (*Mentha piperita* L.) and pine (*Pinus sylvestris* L.) essential oils in human lymphocytes and drosophila melanogaster - Pub Med; 2020.  
Available: <https://pubmed.ncbi.nlm.nih.gov/11313115/>
  11. Heghes SC, Vostinaru O, Rus LM, Mogosan C, Iuga CA, Filip L. Antispasmodic effect of essential oils and their constituents: A Review. *Molecules.* 2019;24(9).  
DOI:10.3390/molecules24091675
  12. Thomson M, Ali M. Garlic [*Allium sativum*]: A review of its potential use as an anti-cancer agent. *Curr Cancer Drug Targets.* 2003;3:67-81.  
DOI:10.2174/1568009033333736
  13. Kendler BS. Garlic (*Allium sativum*) and onion (*Allium cepa*): A review of their relationship to cardiovascular disease. *Prev Med.* 1987;16(5):670-685.  
DOI: 10.1016/0091-7435(87)90050-8
  14. Lazarević JS, Dordević AS, Zlatković B, Radulović N, Palić RM. Chemical composition and antioxidant and antimicrobial activities of essential oil of *Allium sphaerocephalon* L. subsp. *sphaerocephalon* (Liliaceae) inflorescences. *J Sci Food Agric.* 2011;91:322-329.
  15. Silva N, Júnior A. Biological properties of medicinal plants: A review of their antimicrobial activity. *J Venom Anim Toxins Trop Dis.* 2009;16:402-413.  
DOI: 10.1590/S1678-91992010000300006
  16. Paoli S de, Giani TS, Presta GA, et al. Effects of clove (*Caryophyllus aromaticus* L.) on the labeling of blood constituents with technetium-99m and on the morphology of red blood cells. *Braz Arch Biol Technol.* 2007;50(SPE):175-182.  
DOI:10.1590/S1516-89132007000600022
  17. Politeo O, Jukic M, Milos M. Comparison of chemical composition and antioxidant activity of glycosidically bound and free volatiles from clove (*eugenia Caryophyllata* Thunb.). *J Food Biochem.* 2010;34(1):129-141.  
DOI: <https://doi.org/10.1111/j.1745-4514.2009.00269.x>
  18. Koba K, Nenonene AY, Raynaud C, Chaumont J-P, Sanda K. Antibacterial Activities of the buds essential oil of *Syzygium aromaticum* (L.) Merr. & Perry from Togo. *J Biol Act Prod Nat.* 2011; 1(1):42-51.  
DOI:10.1080/22311866.2011.10719072
  19. Machado M, Dinis AM, Salgueiro L, Custódio JBA, Cavaleiro C, Sousa MC. Anti-Giardia activity of *Syzygium aromaticum* essential oil and eugenol: Effects on growth, viability, adherence and ultrastructure. *Exp Parasitol.* 2011;127(4): 732-739.  
DOI:10.1016/j.exppara.2011.01.011
  20. Wang Y, Ocariz J, Hammersand J, et al. Determination of cinnamaldehyde in cinnamon by SPME-GC-MS. An Instrumental Analysis Experiment. *J Chem Educ.* 2008;85.  
DOI: 10.1021/ed085p957
  21. Unlu M, Ergene E, Unlu GV, Zeytinoglu HS, Vural N. Composition, antimicrobial activity and in vitro cytotoxicity of essential oil from *Cinnamomum zeylanicum* Blume (Lauraceae). *Food Chem Toxicol Int J Publ Br Ind Biol Res Assoc.* 2010;48(11):3274-3280.  
DOI: 10.1016/j.fct.2010.09.001
  22. Brezáni V, Karel Š. Secondary metabolites isolated from the genus *Eucalyptus*. *Curr Trends Med Chem.* 2013;7:65-95.
  23. Ben-Arye E, Dudai N, Eini A, Torem M, Schiff E, Rakover Y. Treatment of upper respiratory tract infections in primary care: A randomized study using aromatic herbs.



- Evid-Based Complement Altern Med ECAM. 2011;2011:690346.  
DOI: 10.1155/2011/690346
24. Ahmed SBH, Sghaier RM, Guesmi F, et al. Evaluation of antileishmanial, cytotoxic and antioxidant activities of essential oils extracted from plants issued from the leishmaniasis-endemic region of Sned (Tunisia). Nat Prod Res. 2011;25(12): 1195-1201.  
DOI: 10.1080/14786419.2010.534097
  25. Bandoni A, Van Baren C, Eguaras M, et al. Antimicrobial and miticide activities of eucalyptus globulus essential oils obtained from different argentine regions. Span J Agric Res ISSN 1695-971X No 3 2010 Pags 642-650. 2010;8.  
DOI: 10.5424/sjar/2010083-1260
  26. Verma R, Rahman L ur, Rk V, Chauhan A, Yadav A, AS. Essential oil composition of menthol mint (*Mentha arvensis* L.) and Peppermint (*Mentha piperita* L.) cultivars at different stages of plant growth from kumaon region of Western Himalaya. Open Access J Med Aromat Plants. 2010; 1:13-18.
  27. Sousa D. Analgesic-like activity of essential oils constituents. Mol Basel Switz. 2011;16:2233-2252.  
DOI: 10.3390/molecules16032233
  28. Kumar P, Mishra S, Malik A, Satya S. Insecticidal properties of *Mentha species*: A review. Ind Crops Prod. 2011;34:802-817.  
DOI: 10.1016/j.indcrop.2011.02.019
  29. Sabzghabae AM, Nili F, Ghannadi A, Eizadi-Mood N, Anvari M. Role of menthol in treatment of candidial napkin dermatitis. World J Pediatr. 2011;7(2):167-170.  
DOI: 10.1007/s12519-011-0253-0
  30. Singh R, Shushni MAM, Belkheir A. Antibacterial and antioxidant activities of *Mentha piperita* L. Arab J Chem. 2015;8(3):322-328.  
DOI: 10.1016/j.arabjc.2011.01.019
  31. Hajhashemi V, Ghannadi A, Sharif B. Anti-inflammatory and analgesic properties of the leaf extracts and essential oil of *Lavandula angustifolia* Mill. J Ethnopharmacol. 2003;89:67-71.  
DOI: 10.1016/S0378-8741(03)00234-4
  32. Lee Y-L, Wu Y, Tsang HWH, Leung AY, Cheung W m. A Systematic review on the anxiolytic effects of aromatherapy in people with anxiety symptoms. J Altern Complement Med. 2011;17(2):101-108.  
DOI: 10.1089/acm.2009.0277
  33. Woronuk G, Demissie Z, Rheault M, Mahmoud S. Biosynthesis and therapeutic properties of lavandula essential oil constituents. Planta Med. 2011;77(01):7-15.  
DOI: 10.1055/s-0030-1250136
  34. Hart P, Brand C, Carson C, Riley T, Prager R, Finlay-Jones J. Terpinen-4-ol, the main component of the essential oil of *Melaleuca alternifolia* (tea tree oil), suppresses inflammatory mediator production by activated human monocytes. Inflamm Res - INFLAMM Res. 2000;49:619-626.  
DOI: 10.1007/s000110050639
  35. Vuuren SFV, Suliman S, Viljoen AM. The antimicrobial activity of four commercial essential oils in combination with conventional antimicrobials. Lett Appl Microbiol. 2009;48(4):440-446.  
DOI: <https://doi.org/10.1111/j.1472-765X.2008.02548.x>
  36. Satil F, Selvi S, Polat R. Ethnic uses of pine resin production from *Pinus brutia* by native people on the Kazdağ Mountain (Mt. Ida) in Western Turkey. J Food Agric Environ. 2011;SCI:1059-1063.
  37. Kizilarslan Hancer C, Sevgi E. Ethnobotanical uses of genus *Pinus* L. (Pinaceae) in Turkey. Indian J Tradit Knowl. 2013;12.
  38. Badal S. Pharmacognosy: Fundamentals, Applications and Strategy.; 2016.
  39. Bagci E, Hayta S, Dogan G. Chemical composition of essential oils from bark and leaves of *Pinus brutia* ten from Turkey. Asian J Chem. 2011;23:2782-2784.
  40. Shinde UA, Phadke AS, Nair AM, Mungantiwar AA, Dikshit VJ, Saraf MN. Membrane stabilizing activity — a possible mechanism of action for the anti-inflammatory activity of Cedrus deodara wood oil. Fitoterapia. 1999;70(3):251-257.  
DOI: 10.1016/S0367-326X(99)00030-1
  41. Chatterjee P, Chandra S, Dey P, Bhattacharya S. Evaluation of anti-inflammatory effects of green tea and black tea: A comparative in vitro study. J Adv Pharm Technol Res. 2012;3(2):136-138.  
DOI: 10.4103/2231-4040.97298
  42. Kumari C, Yasmin N, Hussain M, Babu MB. In vitro anti-inflammatory and anti-arthritic property of *Rhizopora mucronata* leaves. Int J Pharma Sci Res. 2015;6.
  43. Loizzo M, Saab A, Tundis R, et al. Chemical composition and antimicrobial activity of essential oils from *Pinus brutia*

- (calabrian pine) growing in Lebanon. Chem Nat Compd. 2008;44:784-786.  
DOI: 10.1007/s10600-009-9167-7
44. Riahi L, Chograni H, Ziadi S, Zoghlami N, Mliki A. Essential oils of *Pinus brutia* and *Cupressus sempervirens* from Tunisia: chemical composition and antioxidant activity; 2012.
45. Ge W, Li D, Gao Y, Cao X. The Roles of Lysosomes in Inflammation and Autoimmune Diseases. Int Rev Immunol. 2014;34.  
DOI: 10.3109/08830185.2014.936587
46. Elisha IL, Dzoyem J, Mcgaw L, Botha F, Eloff J. The anti-arthritic, anti-inflammatory, antioxidant activity and relationships with total phenolics and total flavonoids of nine South African plants used traditionally to treat arthritis. BMC Complement Altern Med. 2016;16:307.  
DOI: 10.1186/s12906-016-1301-z
47. Miguel MG. Antioxidant and anti-inflammatory activities of essential oils: A Short Review. Molecules. 2010;15(12): 9252-9287.  
DOI: 10.3390/molecules15129252
48. Süntar I, Tumen I, Ustün O, Keles H. Appraisal on the wound healing and anti-inflammatory activities of the essential oils obtained from the cones and needles of *Pinus* species by in vivo and in vitro experimental models. J Ethnopharmacol. 2011;139:533-540.  
DOI: 10.1016/j.jep.2011.11.045
49. Held S, Schieberle P, Somoza V. Characterization of alpha-terpineol as an anti-inflammatory component of orange juice by in vitro studies using oral buccal cells. J Agric Food Chem. 2007;55(20): 8040-8046.  
DOI: 10.1021/jf071691m

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